

## A Meta-Analysis of Essential Amino Acid Requirements of Fish

Salze Guillaume\*, Quinton Margaret, and Bureau Dominique P.

Department of Animal and Poultry Science, University of Guelph, Guelph, ON, Canada,  
N1G 2W1, Current E-mail: [gSalze@auburn.edu](mailto:gsalze@auburn.edu)

---

### Abstract

There are wide variations in the published estimates of essential amino acid (EAA) requirements. Variations are thought to originate from different choices in mode of expression, response variable, and mathematical model. Here we conduct a meta-analysis of the growth-based, dose-response trials of 10 EAA in 22 teleost species: 249 studies were reviewed. Published data were entered in a spreadsheet and re-calculated across in a standard and systematic manner to allow comparisons. The considered unit of requirement were percentage of dry diet, g of EAA per MJ of digestible energy (DE), and g of ingested EAA per kg of metabolic body weight (MBW) per day. Response variables included growth in g per kg MBW per day and thermal-unit growth coefficient (TGC). Four mathematical models were also compared: broken-lime model (BLM), quadratic model (QM), broken-quadratic model (BQM), and saturation kinetic model (SKM). Results first indicate important differences in study quality, as 54% of the reviewed papers were excluded from the meta-analysis, often times because of poor growth or missing information. Additionally, the final dataset was greatly fragmented: 31% of the studied concerned rainbow trout, and lysine was the focus of 29% of all studies, leaving some species and EAA poorly covered. Comparisons of the requirement estimates show important variations between studies, even within species. With such variability there was no difference in requirement estimates calculated with different response variables. Similarly, this variability was not different between the three modes of expression, nor was it between mathematical models. However, there were significant effects of experimental design on the quality of fit of the models. Specifically, experiments that failed to produce a clear, plateauing dose-response curve had greatly increased probability to yielding absurd results (e.g. negative requirement). Finally, the present study emphasizes the critical need of a global, standard and systematic system to report and capture results from nutrition trials.

Keywords: Meta-analysis, amino acid, requirement

## 1 Introduction

Aquafeed formulations are constantly refined, updated, and adjusted to reflect progress in nutrition science, as well as the variability in ingredients price and availability. Feeds must remain cost-effective and deliver nutrients efficiently to the animal in order to promote growth and health while minimizing nutrient discharge in the effluent. As a result, fish feed are increasingly formulated at the nutrient level, as is commonly done in other livestock, e.g., poultry. Naturally, the first prerequisite to this goal is a precise knowledge of essential amino acid (EAA) requirement.

However, a number of reviews have identified significant gaps in our understanding of EAA requirements of teleosts and highlighted the great variability in estimates of EAA requirements amongst and within species (Bureau and Encarnaçao, 2006; Cowey, 1994; Lall and Anderson, 2005; NRC, 1993, 2011; Wilson, 1989). For example, in rainbow trout *Oncorhynchus mykiss* lysine requirement range from 1.3 to 2.9% of the diet (3.7-7.3% of crude protein, CP), and arginine requirement estimates span from 1.4 to 2.2% of the diet. Genetic variations likely account for but a fraction of such different estimates. Rather, reasons explaining these discrepancies can be found in different methodological approaches. Calculation of a requirement estimate requires choosing a mode of expression, a response variable, and a mathematical model to describe their relationship. Modes of expression include percentage of the diet or crude protein, or in g/MJ digestible energy. The choice of mode of expression is a matter of much debate, and reflects the various assumptions authors make when considering what affects the requirements or not (Bureau and Encarnaçao, 2006; Bureau, 2008). Because these assumptions contradict each other, the same diet can be deemed deficient or not depending on the mode of expression of EAA requirement. Similarly, the choice of the response variable likely influences the estimates of requirements, as it introduces another level of variability between experiments. Responses commonly used in this context include expressions of weight gain (absolute weight gain, specific growth rate, percentage increase, thermal-unit growth coefficient, etc.), feed efficiency, nitrogen retention, or, more rarely, blood EAA concentration. Most studies consider growth as the primary response, but they do not all use the same starting weight in a given species. This is also problematic, since biological processes governing growth in fish are quite dynamic and

change significantly during the production cycle (Dumas *et al.* 2007), and EAA requirements likely change as well. Mathematical models complete the triptych of the analytical aspect of nutrient requirement estimation. Different models have been used to describe the fish biological response, which typically follows the law of “diminishing return”. The broken-line model (BLM) has been heavily used for the past two decades. However, when compared to other non-linear models, the BLM consistently returns lower estimates (Hernandez-Llamas, 2009). In contrast, Shearer (2000) noted that non-linear models could yield requirement estimates up to five times the published requirement.

Clearly, there is a dire need to re-visit fish EAA requirements. Thus, the present study first extensively reviewed the existing literature on EAA requirements in fish and systematically standardized the relevant data. The resulting dataset was then analyzed using various modes of expression, responses and mathematical models to re-evaluate the EAA requirement in these commercially important fish species.

## 2 Methods and calculations

### 2.1 Search of the literature and selection criteria

An extensive search of the EAA requirement literature was conducted using online databases, peer-reviewed journals, and books. At this point, the sole selection criterion was to only keep studies on commercially-relevant species (e.g., salmonidae, cyprinidae, cichlidae), resulting in 286 articles. There was among these a great variety of objectives, experimental designs, and analytical methodologies employed. Therefore, additional selection criteria were applied to the dataset to eliminate studies unsuitable to the present meta-analysis.

Since growth was by far the most studied type of response, only studies based on growth trials in response to graded dietary levels of a unique AA were kept. Thus experiments based on the measurement of plasma AA levels, or on the ideal protein theory, were not included (37 studies out of the initial 286).

Other selection criteria were as follow: husbandry practices must be adequate for the species of interest, including water quality. Information on temperature and experiment duration must be provided. All experimental diets must be adequately characterized and meet all other

known nutritional requirements. Studies must use five or more experimental diets in order to ensure the accuracy of the non-linear regressions analyses. Data, especially diet composition, must be reported on a dry-matter basis, or enough information must be provided to compute dry-matter composition of the diets. Studies where mortality was high and uncorrelated with EAA graded levels were not included. Finally, initial and final individual body weight, as well as feed intake, must be reported, or enough information must be provided to calculate them.

## 2.2 Variables and models combinations

The meta-analysis was conducted as a 3x2x4 array: three factor variables (percentage of EAA of interest in the dry diet, amount of EAA per MJ of digestible energy, and the amount of the EAA of interest ingested during the experiment), and two response variables (weight gain per kg of metabolic body weight and thermal-unit growth coefficient, TGC). Together, they form six pairs of variable. Finally, four regression models (broken-line, quadratic, broken-quadratic, and saturation-kinetic models) were used to estimate the EAA requirements for each of the variable pair and for each selected study.

### 2.2.1 Calculated factor and response variables

Weight gain, on a metabolic body weight-basis ( $WG_{MBW}$ , in g/kgMBW/d)

$$WG_{MBW} = \frac{FBW - IBW}{\left[ \left( \frac{FBW^{1/3} + IBW^{1/3}}{2} \right)^3 \times 10^{-3} \right]^{0.8}} \times d$$

Thermal-unit growth coefficient (TGC, dimensionless)

$$TGC = 100 \times \frac{FBW^{1/3} - IBW^{1/3}}{t \times d}$$

AA dietary level to digestible energy (AA<sub>DE</sub>, in g/MJ<sub>DE</sub>)

$$AA_{DE} = \frac{AA_D \times 10}{GE_D \times ADC_{GE}}$$

AA ingested (AA<sub>I</sub>, in g)

$$AA_I = \frac{AA_D}{100} \times FI$$

Where IBW and FBW are the individual initial and final body weight (in g) respectively; d is the duration of the experiment (in days), t is the average temperature during the experiment (in °C); AA<sub>D</sub> is the diet content of the amino acid of interest (% dry matter); GE<sub>D</sub> is the gross energy content of the diet (in MJ.kg<sup>-1</sup>); ADC<sub>GE</sub> is the apparent digestibility coefficient of gross energy (%); FI is the feed intake during the experiment (in g, dry matter basis).

### 2.2.2 Models

The broken line model (BLM) is defined as follow:

$$Y = a_1X + b_1 + (a_2 - a_1)\delta X + (b_2 - b_1)\delta + \varepsilon$$

$\delta=0$  if  $X \leq X_{bp}$ , or  $\delta=1$  if  $X \geq X_{bp}$

Where Y and X are the response and independent variables, respectively; X<sub>bp</sub> is the abscissa of the breaking point thus defining the AA requirement level; b<sub>1</sub> and b<sub>2</sub> are the intercepts, and a<sub>1</sub> and a<sub>2</sub> the slopes, of the 2 lines, respectively. Note that a<sub>2</sub> was not constrained to zero. The quadratic model (QM) is defined as follow:

$$Y = aX^2 + bX + c + \varepsilon$$

Where Y and X are the response and independent variables, respectively, while a, b, and c are equation parameters. The amino acid requirement is the abscissa of the smaller root that solve the parabola's equation for Y=95% of the maximum response.

The following equation defines the broken-quadratic model (BQM):

$$Y = aX^2 + bX + c + a(X_{jp}^2 - X^2)\delta + b(X_{jp} - X)\delta + \varepsilon$$

$\delta=0$  if  $X \leq X_{jp}$ , or  $\delta=1$  if  $X \geq X_{jp}$

Where  $Y$  and  $X$  are the response and independent variables, respectively, and  $a$ ,  $b$ , and  $c$  are equation parameters.  $X_{jp}$  is the abscissa of the junction point thus defining the AA requirement level. Note that since  $X_{jp}$  is a constant, the curve becomes a line of slope zero for amino acid levels above the estimated requirement.

Finally, the saturation-kinetic model (SKM) is defined as follow:

$$Y = \frac{b \times c + aX^d}{c + X^d} + \varepsilon$$

Where  $Y$  and  $X$  are the response and independent variables, respectively;  $a$  is the asymptote to the maximum response;  $b$  is the intercept;  $c$  is a coefficient;  $d$  is a kinetic order. The amino acid requirement is defined as the abscissa where the response variable reaches 95% of its predicted maximum (i.e. 95% of  $a$ ).

Estimations of EAA requirements were computed for each of the independent/dependent variable couples and for each of the 4 models. Outputs were classified into the following categories: “no fit” indicates that the model failed to converge, and therefore no estimate of requirement was generated. “Within” means that the model converged and the estimated requirement fell within the range of experimental EAA levels tested in a given study. “Outside” signifies that the model successfully converged, but the estimate requirement fell outside of the EAA range of the experimental diets. “No break”, for the BLM and BQM, indicates that only one of the two curves calculated by the model fitted, hence no breaking point could be determined. Finally, the quadratic-based models (BQM and QM) sometimes converged to a positive  $a$ , thus causing the model to have a minimum instead of a maximum. This behaviour was noted as “minimum”.

### 2.3 Statistical analysis

Models were programmed and run using SAS 9.4 (SAS Institute). Qualities of fit and coefficient of variation of requirement estimates were tested by logistic tests and a two-way ANOVA (variable pairs, models), respectively.

### 3 Results and discussion

#### 3.1 Screening results

A total of 348 peer-reviewed articles spanning 52 years (1964 to 2016) were found during the literature search, out of which 48 were not growth-based trials – e.g. based on ideal-protein formulations, or on changes in plasma AA levels, review articles – hence could not be included in the meta-analysis. However, other selection criteria rejected over half (52%) of the 300 remaining studies from the meta-analysis (Figure 1). The main reasons for rejection included poor growth, missing information (e.g. dry matter data, feed intake), or too few graded levels (

**Table 1).** The final dataset (Table 2) 145 studies which covered 37 species of fish, and 11 EAA. About 72% of the studies used 6 or 7 graded levels of the EAA of interest. One study used 24 graded levels. The variables most commonly used for the estimation of the requirement are the EAA content of the diet (% dry matter) and weight gain (as SGR or % of IBW) as the independent and response variable, respectively. The most commonly used models are the BLM and QM.

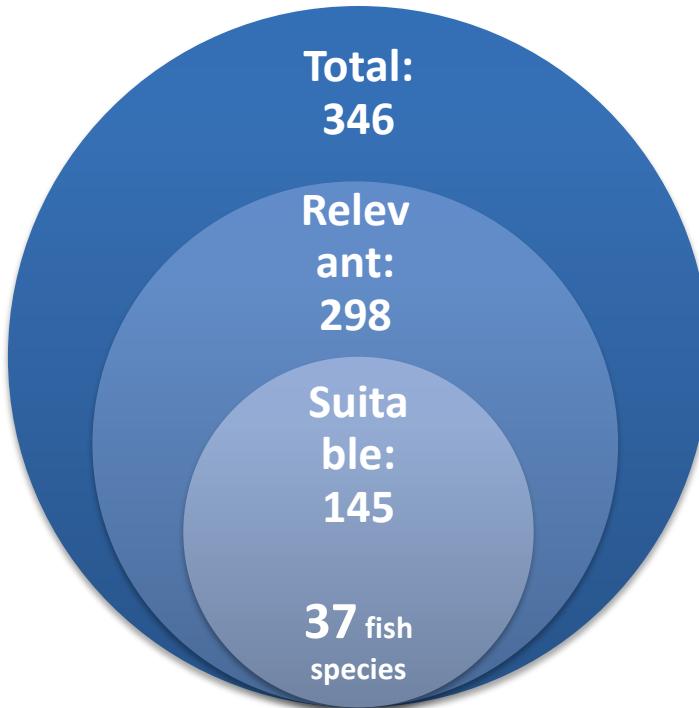


Figure 1: Diagram illustrating the screening of studies, from the total number found to the number of suitable studies that constituted our working data set

Table 1: Study rejection

<b>Reason for rejection</b>	<b>% of rejected studies</b>
<b>Restricted feeding</b>	3%
<b>Poor growth</b>	34%
<b>Too few dietary levels</b>	10%
<b>Lack of intake data</b>	44%
<b>Lack of dry matter data</b>	17%
<b>Lack of raw growth data</b>	2%

The number of excluded studies is striking. It highlights the uneven quality of the research effort in this area. It also exacerbated the fragmentation of the final dataset, both in terms of species and EAA. For instance, 25% of the studies focused on rainbow trout, whereas studies on other species such as European seabass, Japanese flounder, or Nile tilapia each represent 3% or less of the dataset. Similarly, lysine is by far the most-studied EAA (27% of the studies), while only five studies (3% of the dataset, Figure 2) describe the requirement of

phenylalanine. Moreover, these five studies were performed on five different species. Such fragmentation constitutes a major challenge for the understanding and definition of EAA requirements, *let al* one their rationalization.

### 3.2 Estimates of EAA requirements

There were least two studies in 13 of the possible combinations of species and EAA (Table 3). These 13 combinations reflect the most-studied EAA (arginine, isoleucine, lysine, methionine, threonine, and tryptophan) and species (channel catfish, hybrid striped bass, rainbow trout, and Chinook, Coho and Atlantic salmons). Estimations are generally well in agreement with those reported by the NRC (2011): e.g., in % of diet dry matter, 1.66% arginine in rainbow trout, 1.53% lysine in channel catfish, or 0.61% methionine in hybrid striped bass.

**Table 2: Summary of studies included in the final dataset**

Studied EAA	Species	# of diet	EAA range in diets (% DM)	Av. water temp.	Av. IBW (g)	$\frac{maxFBW}{IBW}$	$\frac{minFBW}{maxFBW}$	Exp. Duration (days)	Reference
Arg	<i>Clarias hybrid</i>	6	1.06-2.28	26.0	6.71	6.23	0.49	28	(Singh and Khan, 2007)
	<i>Ictalurus punctatus</i>	8	0.20-1.60	26.7	200	2.21	0.51	56	(Robinson <i>et al.</i> 1981)
	<i>Heteropneustes fossilis</i>	6	0.85-2.13	23.5	4.83	3.77	0.49	56	(Ahmed, 2013)
	<i>Labeo rohita</i>	6	0.47-1.77	28.0	0.62	5.00	0.37	56	(Abidi and Khan, 2009)
	<i>Cirrhinus mrigala</i>	6	1.00-2.25	26.5	0.61	4.21	0.55	56	(Ahmed and Khan, 2004b)
	<i>Ctenopharyngodon idella</i>	6	0.93-2.41	27.8	3.85	5.53	0.50	70	(Gao <i>et al.</i> 2015)
	<i>Catla catla</i>	6	0.98-2.23	27.9	0.60	12.41	0.46	84	(Zehra and Khan, 2013a)
	<i>Micropterus salmoides</i>	6	1.70-3.01	28.0	24.93	2.77	0.88	56	(Zhou <i>et al.</i> 2012)
	<i>Oncorhynchus kisutch</i>	6	1.20-3.60	10.0	2.90	3.30	0.65	70	(Klein and Halver, 1970)
	<i>Oncorhynchus tshawytscha</i>	6	1.60-3.60	10.0	3.48	2.89	0.79	70	(Klein and Halver, 1970)
	<i>Oncorhynchus kisutch</i>	6	0.00-5.27	15.0	0.91	5.90	0.87	56	(Luzzana <i>et al.</i> 1998)
	<i>Oncorhynchus mykiss</i>	12	0.5-2.38	17.0	50.67	3.54	0.54	51	(Rodehutscord <i>et al.</i> 1995a)
	<i>Oncorhynchus mykiss</i>	8	0.47-2.50	15.0	12.60	3.35	0.64	42	(Kim <i>et al.</i> 1992b)
	<i>Oncorhynchus mykiss</i>	6	0.69-3.83	15.0	2.50	14.08	0.65	84	(Cho <i>et al.</i> 1992)
	<i>Oncorhynchus mykiss</i>	7	0.69-3.00	15.0	1.30	19.46	0.72	84	(Cho <i>et al.</i> 1992)
	<i>Oncorhynchus mykiss</i>	5	0.80-2.80	10.0	7.30	4.08	0.60	84	(Walton <i>et al.</i> 1986)
	<i>Oncorhynchus mykiss</i>	5	1.72-3.61	17.0	9.30	9.46	0.74	84	(Fournier <i>et al.</i> 2003)
	<i>Salmo salar</i>	5	1.13-2.86	6.5	385	1.50	0.91	126	(Berge <i>et al.</i> 1997)
	<i>Salmo salar</i>	5	1.10-3.20	15.0	111.9	5.00	0.87	98	(Lall <i>et al.</i> 1994)
	<i>Psetta maxima</i>	5	1.79-4.19	17.0	7.40	8.03	0.43	84	(Fournier <i>et al.</i> 2003)

	<i>Sparus macrocephalus</i>	6	1.85-3.46	28.0	10.51	4.87	0.84	56	(Zhou et al. 2010b)
	<i>Lates calcarifer</i>	6	0.73-3.73	29.0	2.56	7.96	0.64	84	(Murillo-Gurrea et al. 2001)
	<i>Rachycentron canadum</i>	6	1.76-3.75	30.5	3.38	13.11	0.76	63	(Ren et al. 2014)
	<i>Paralichthys olivaceus</i>	6	1.25-3.25	20.6	1.85	4.25	0.53	40	(Alam et al. 2002)
His	<i>Ictalurus punctatus</i>	9	0.10-0.80	26.7	200	3.31	0.39	56	(Wilson et al. 1980)
	<i>Cirrhinus mrigala</i>	6	0.25-1.50	27.5	0.61	3.95	0.60	56	(Ahmed and Khan, 2005b)
	<i>Catla catla</i>	6	0.25-0.96	28.1	0.65	14.33	0.03	84	(Zehra and Khan, 2016c)
	<i>Heteropneustes fossilis</i>	6	0.90-1.95	27.9	5.27	14.33	0.08	84	(Khan and Abidi, 2014)
	<i>Oncorhynchus mykiss</i>	12	0.26-1.35	17.0	40.25	4.22	0.71	53	(Rodehutscord et al. 1997)
	<i>Oncorhynchus keta</i>	7	0.13-1.92	14.8	1.58	2.46	0.64	28	(Akiyama et al. 1985)
	<i>Oncorhynchus kisutch</i>	5	0.50-1.30	10.0	3.17	3.00	0.88	70	(Klein and Halver, 1970)
Ile	<i>Ictalurus punctatus</i>	7	0.30-1.30	26.7	200	2.95	0.47	56	(Wilson et al. 1980)
	<i>Labeo rohita</i>	6	0.75-1.99	26.7	0.62	4.46	0.62	56	(Khan and Abidi, 2007b)
	<i>Cirrhinus mrigala</i>	6	0.50-1.75	27.7	0.61	4.18	0.52	56	(Ahmed and Khan, 2006)
	<i>Catla catla</i>	6	0.51-1.73	27.8	0.61	11.69	0.43	84	(Zehra and Khan, 2013c)
	<i>Oncorhynchus mykiss</i>	12	0.50-1.53	16.0	45.65	3.43	0.55	59	(Rodehutscord et al. 1997)
	<i>Salvelinus namaycush</i>	7	0.54-1.26	8.3	3.20	4.00	0.94	84	(Hughes et al. 1983)
	<i>Oncorhynchus tshawytscha</i>	6	0.50-1.50	10.0	0.82	4.26	0.68	70	(Chance et al. 1964)
	<i>Oncorhynchus tshawytscha</i>	6	0.50-1.50	10.0	0.79	4.14	0.61	70	(Chance et al. 1964)
Leu	<i>Ictalurus punctatus</i>	8	0.60-2.00	26.7	200	2.87	0.76	56	(Wilson et al. 1980)

<i>Labeo rohita</i>	6	0.74-1.99	26.7	0.59	4.00	0.46	56	(Abidi and Khan, 2007)
<i>Cirrhinus mrigala</i>	6	0.75-2.00	27.7	0.64	4.08	0.55	56	(Ahmed and Khan, 2006)
<i>Oreochromis niloticus</i>	6	0.53-1.81	29.0	1.94	10.77	0.70	56	(Gan <i>et al.</i> 2016)
<i>Salvelinus namaycush</i>	7	0.96-2.24	8.3	3.20	4.03	0.84	84	(Hughes <i>et al.</i> 1983)
<i>Oncorhynchus tshawytscha</i>	6	1.00-3.10	10.0	2.49	2.11	0.60	56	(Chance <i>et al.</i> 1964)
<i>Oncorhynchus mykiss</i>	12	1.00-4.20	15.7	49.33	3.77	0.66	53	(Rodehutscord <i>et al.</i> 1997)
<i>Lateolabrax japonicus</i>	6	0.90-3.88	29.0	8.00	4.90	0.85	56	(Li <i>et al.</i> 2015)
Lys	<i>Hybrid striped bass</i>	6	1.14-2.37	24.5	7.0	6.76	0.78	63 (Keembiyehetty and Gatlin, 1992)
	<i>Hybrid striped bass</i>	8	1.20-2.60	28.0	5.5	10.46	0.85	77 (Griffin <i>et al.</i> 1992)
	<i>Ictalurus punctatus</i>	6	0.75-2.00	26.7	202	2.50	0.66	44 (Wilson <i>et al.</i> 1977)
	<i>Ictalurus punctatus</i>	6	0.70-2.00	26.7	201	2.69	0.51	56 (Robinson <i>et al.</i> 1980b)
	<i>Pelteobagrus fulvidraco</i>	6	1.73-4.19	28.0	1.50	8.00	0.70	56 (Cao <i>et al.</i> 2012)
	<i>Lepomis macrochirus</i>	7	1.19-3.29	22.3	26.93	1.50	0.83	56 (Masagounder <i>et al.</i> 2011)
	<i>Ctenopharyngodon idella</i>	6	0.71-3.16	28.7	3.15	4.49	0.64	90 (Wang <i>et al.</i> 2005)
	<i>Cirrhinus mrigala</i>	6	1.48-2.78	26.5	0.66	3.95	0.59	40 (Ahmed and Khan, 2004a)
	<i>Catla catla</i>	6	1.22-2.47	26.8	0.59	12.32	0.45	84 (Zehra and Khan, 2013b)
	<i>Piaractus mesopotamicus</i>	6	0.82-2.10	29.6	6.67	7.08	0.61	90 (Abimorad <i>et al.</i> 2010)
	<i>Oncorhynchus keta</i>	8	0.38-4.00	14.8	1.27	3.66	0.57	28 (Akiyama <i>et al.</i> 1985)

								(Rodehutscord 1997)	<i>et</i> <i>al.</i>
<i>Oncorhynchus mykiss</i>	24	0.45-5.80	15.7	50.9	3.65	0.32	55		
<i>Oncorhynchus mykiss</i>	6	1.60-2.42	14.5	14.9	6.00	0.83	56	(Cheng <i>et al.</i> , 2003)	
<i>Oncorhynchus mykiss</i>	8	1.37-3.66	15.0	5.0	16.38	0.65	84	(Wang <i>et al.</i> , unpublished)	
<i>Oncorhynchus mykiss</i>	6	1.27-2.32	15.0	24.0	7.20	0.69	84	(Encarnacao <i>et al.</i> 2004)	
<i>Oncorhynchus mykiss</i>	6	1.33-2.49	15.0	24.0	7.54	0.75	84	(Encarnacao <i>et al.</i> 2004)	
<i>Oncorhynchus mykiss</i>	8	0.72-1.60	15.0	13.7	3.52	0.65	42	(Kim <i>et al.</i> 1992b)	
<i>Oncorhynchus mykiss</i>	8	0.21-3.01	11.5	17.0	4.61	0.40	71	(Pfeffer <i>et al.</i> 1992)	
<i>Oncorhynchus mykiss</i>	6	1.03-2.02	15.0	0.92	5.15	0.57	42	(Nang Thu <i>et al.</i> 2007)	
<i>Oncorhynchus mykiss</i>	6	1.00-2.03	15.0	0.92	3.64	0.58	42	(Nang Thu <i>et al.</i> 2007)	
<i>Oncorhynchus mykiss</i>	6	1.06-2.05	15.0	0.92	5.18	0.54	42	(Nang Thu <i>et al.</i> 2007)	
<i>Oncorhynchus mykiss</i>	6	1.03-2.02	15.0	0.92	5.32	0.49	30	(Nang Thu <i>et al.</i> 2009)	
<i>Oncorhynchus mykiss</i>	6	0.63-1.43	15.0	0.92	3.79	0.48	30	(Nang Thu <i>et al.</i> 2009)	
<i>Oncorhynchus mykiss</i>	6	1.03-2.02	15.0	0.92	4.51	0.47	30	(Nang Thu <i>et al.</i> 2009)	
<i>Oncorhynchus mykiss</i>	7	1.04-2.60	15.0	5.19	10.87	0.50	84	(Walton <i>et al.</i> 1984b)	
<i>Salmo salar</i>	7	1.24-2.94	15.0	4.72	2.71	0.61	70	(Anderson <i>et al.</i> 1993)	
<i>Salmo salar</i>	12	1.20-2.38	15.2	42.12	2.63	0.64	76	(Hauler <i>et al.</i> 2007)	
<i>Salmo salar</i>	7	1.26-4.15	8.0	643	1.60	0.89	85	(Espe <i>et al.</i> 2007)	
<i>Lates calcarifer</i>	6	1.00-2.50	27.0	13.12	4.58	0.70	84	(Murillo-Gurrea <i>et al.</i> 2001)	
<i>Sparus macrocephalus</i>	6	2.08-4.05	18.0	9.12	4.14	0.85	56	(Zhou <i>et al.</i> 2009, 2010a)	
<i>Pagrus major</i>	6	1.19-3.44	22.5	1.7	8.06	0.58	56	(Forster and Ogata, 1998)	
<i>Paralichthys olivaceus</i>	6	1.04-3.16	19.5	3.0	5.33	0.45	63	(Forster and Ogata, 1998)	

<i>Psetta maxima</i>	7	1.20-3.11	18.0	18.1	1.82	0.88	56	(Peres and Oliva-Teles, 2008)
<i>Rachycentron canadum</i>	6	1.15-3.25	28.5	1.26	19.09	0.66	56	(Zhou et al. 2007)
<i>Sciaenops ocellatus</i>	7	1.00-2.50	27.0	6.72	4.57	0.68	56	(Craig and Gatlin, 1992)
<i>Dicentrarchus labrax</i>	5	1.33-2.72	25.5	0.85	7.73	0.65	70	(Tibaldi and Lanari, 1991)
Met	<i>Hybrid striped bass</i>	6	0.38-1.60	24.0	4.8	5.12	0.20	56
	<i>Hybrid striped bass</i>	5	0.60-1.00	24.0	8.6	5.38	0.51	56
	<i>Oreochromis niloticus</i>	5	0.56-0.80	26.2	1.31	6.05	0.78	43
	<i>Megalobrama amblycephala</i>	6	0.39-1.54	28.5	3.34	7.33	0.73	63
	<i>Catla catla</i>	6	0.56-1.81	26.9	0.65	14.77	0.39	84
	<i>Ictalurus punctatus</i>	7	0.25-1.75	26.7	202	2.39	0.71	44
	<i>Ictalurus punctatus</i>	7	0.35-1.55	24.8	14.36	2.34	0.85	42
	<i>Myxocyprinus asiaticus</i>	6	0.64-1.89	27.9	1.72	6.10	0.67	56
	<i>Pseudobagrus ussuriensis</i>	6	0.49-2.08	24.0	0.60	6.16	0.72	56
	<i>Oncorhynchus mykiss</i>	12	0.22-1.09	16.8	43.04	3.25	0.43	49
	<i>Oncorhynchus mykiss</i>	12	0.21-1.10	16.8	43.46	3.59	0.42	49
	<i>Oncorhynchus mykiss</i>	7	0.43-2.07	15.0	3.13	15.48	0.80	112
	<i>Oncorhynchus mykiss</i>	6	0.23-0.80	15.0	15.4	3.08	0.67	42
	<i>Salvelinus alpinus</i>	6	0.43-0.91	12.0	20.55	5.04	0.34	112
	<i>Salmo salar</i>	6	0.69-1.29	8.5	495	2.44	0.87	85

	<i>Paralichthys olivaceus</i>	6	0.53-2.03	21.6	2.8	4.45	0.34	40	(Alam et al. 2000)
	<i>Sparus macrocephalus</i>	6	0.75-2.35	28.0	14.22	4.36	0.84	56	(Zhou et al. 2011)
	<i>Sciaenops ocellatus</i>	7	0.35-1.85	27.0	0.9	14.39	0.19	56	(Moon and Gatlin III, 1991)
	<i>Trachinotus ovatus</i>	6	0.86-1.45	29.5	12.40	6.58	0.64	56	(Niu et al. 2013)
	<i>Rachycentron canadum</i>	6	0.61-1.68	29.0	11.62	7.37	0.62	56	(Zhou et al. 2006)
	<i>Lates calcarifer</i>	6	0.62-1.26	26.5	2.61	11.10	0.86	84	(Coloso et al. 1999)
Phe	<i>Ictalurus punctatus</i>	7	0.20-0.80	26.7	199	2.50	0.60	56	(Robinson et al. 1980a)
	<i>Cirrhinus mrigala</i>	6	0.50-1.75	26.2	0.56	3.88	0.54	56	(Ahmed, 2009)
	<i>Labeo rohita</i>	6	0.39-1.64	26.0	0.58	4.14	0.60	56	(Khan and Abidi, 2007a)
	<i>Catla catla</i>	6	0.39-1.62	27.2	0.68	13.10	0.43	84	(Zehra and Khan, 2014)
	<i>Oncorhynchus mykiss</i>	7	0.26-1.75	15.0	12.7	2.91	0.55	42	(Kim, 1993)
Thr	<i>Ictalurus punctatus</i>	7	0.30-1.20	26.7	200	2.79	0.61	56	(Wilson et al. 1978)
	<i>Hybrid striped bass</i>	6	0.49-1.75	26.5	9.8	4.11	0.73	49	(Keembiyehetty and Gatlin, 1997)
	<i>Hybrid striped bass</i>	6	0.49-1.25	26.5	3.0	6.46	0.69	56	(Keembiyehetty and Gatlin, 1997)
	<i>Cirrhinus mrigala</i>	6	1.00-2.25	26.7	0.52	4.05	0.56	56	(Ahmed et al. 2004)
	<i>Labeo rohita</i>	6	0.71-2.15	27.0	0.58	4.14	0.49	56	(Abidi and Khan, 2008)
	<i>Catla catla</i>	6	0.74-1.93	26.9	0.60	14.32	0.40	84	(Zehra and Khan, 2016a)
	<i>Oreochromis niloticus</i>	6	0.71-2.46	28.8	6.00	3.49	0.63	56	(He et al. 2016)
	<i>Oreochromis niloticus</i>	5	0.89-1.54	28.5	563.30	1.47	0.92	28	(Michelato et al. 2016)
	<i>Megalobrama amblycephala</i>	5	0.61-2.49	27.0	3.01	4.30	0.70	63	(Habte-Tsion et al. 2015)
	<i>Oncorhynchus keta</i>	8	0.24-2.80	14.8	1.19	2.55	0.53	28	(Akiyama et al. 1985)
	<i>Oncorhynchus mykiss</i>	11	0.50-2.18	14.7	1.84	3.52	0.50	24	(Bodin et al. 2008)

								(Rodehutscord 1995a) <i>et al.</i>
	<i>Oncorhynchus mykiss</i>	11	0.50-2.10	17.0	50.74	3.25	0.61	51
	<i>Salmo salar</i>	7	0.00-0.75	14.5	0.80	2.90	0.46	36
	<i>Salmo salar</i>	11	0.50-2.18	14.7	0.80	3.00	0.47	36
	<i>Dicentrarchus labrax</i>	6	0.81-2.63	21.1	7.50	2.80	0.89	65
	<i>Sciaenops ocellatus</i>	6	0.49-1.25	27.0	2.80	8.09	0.41	56
Trp	<i>Ictalurus punctatus</i>	6	0.05-0.40	26.7	200	2.87	0.47	56
	<i>Heteropneustes fossilis</i>	8	0.10-0.39	27.9	6.66	10.90	0.29	72
	<i>Heteropneustes fossilis</i>	6	0.04-0.54	22.8	4.45	3.59	0.50	56
	<i>Labeo rohita</i>	6	0.22-0.44	25.8	0.36	6.81	0.31	84
	<i>Cirrhinus mrigala</i>	6	0.06-0.56	27.7	0.62	3.77	0.47	56
	<i>Catla catla</i>	6	0.10-0.34	27.7	0.60	16.44	0.31	84
	<i>Oncorhynchus mykiss</i>	12	0.13-0.56	15.5	49.72	3.54	0.58	64
	<i>Oncorhynchus mykiss</i>	5	0.01-0.43	9.0	5.45	4.60	0.24	112
Val	<i>Oncorhynchus mykiss</i>	6	0.08-0.60	15.0	13.76	3.82	0.36	84
	<i>Ictalurus punctatus</i>	7	0.40-1.60	26.7	201	2.77	0.61	56
	<i>Labeo rohita</i>	6	0.75-2.00	26.0	0.16	2.69	0.70	42
	<i>Cirrhinus mrigala</i>	6	0.75-2.00	27.7	0.62	4.13	0.54	56
	<i>Oncorhynchus mykiss</i>	12	0.62-3.42	15.7	49.36	3.55	0.43	53
	<i>Oncorhynchus tshawytscha</i>	6	0.65-1.90	10.0	0.67	3.90	0.78	70
								(Chance <i>et al.</i> 1964)

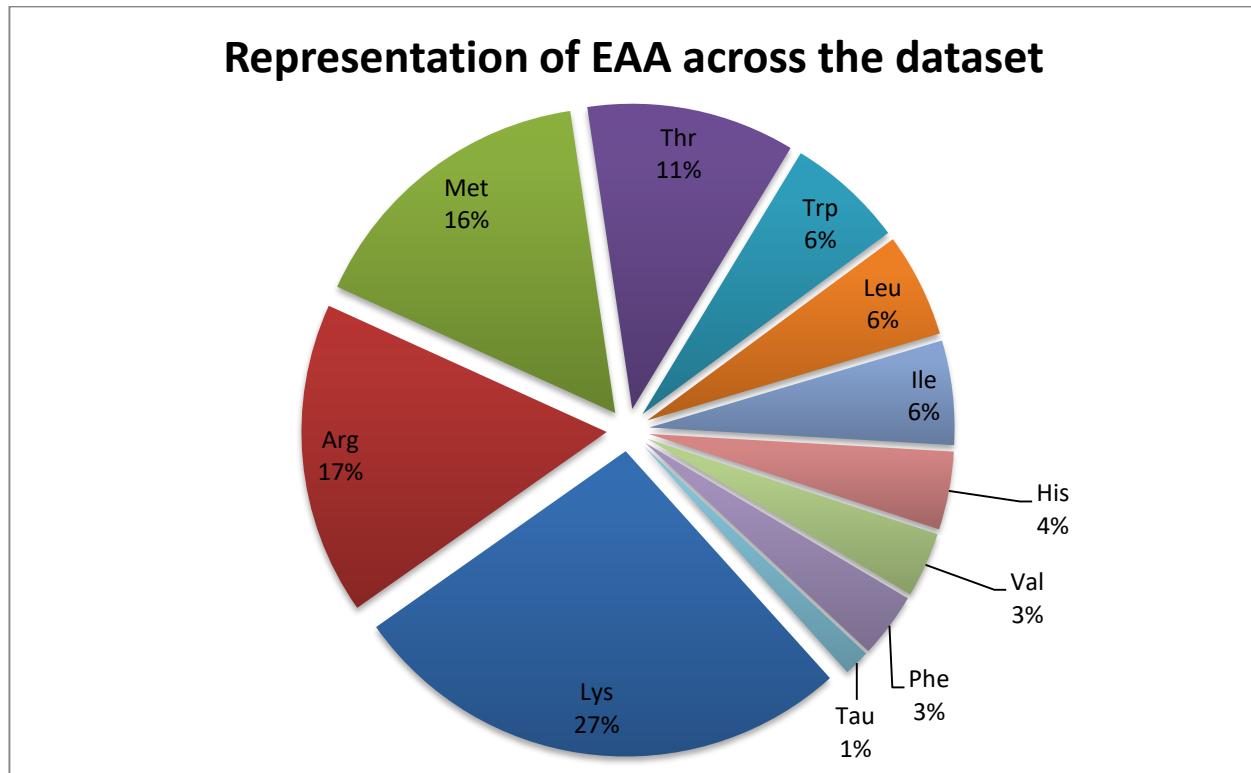


Figure 2: Representation of the eleven EAA in the dataset

In Table 3 we also see the requirements estimated with different variable pairs. While the influence of different response variable can be observed directly, it is meaningless to compare requirements computed with different modes of expression, since they are expressed in different units. Instead, the coefficient of variation (CV) of requirement estimates provides a good indication of the estimate consistency between variable pairs. Regardless of the species and EAA, CVs hover around 22% (median is 15% across the full dataset), which is comparable to the 34% found by Hauler and Carter (2001), but far less than the 100%+ described by Shearer (2000). Bodin *et al.* (2009) found that the choice of response variable impacts the relative lysine requirement in rainbow trout, but not the absolute (i.e. lysine intake) requirement. In our dataset, absolute requirement estimates are indeed numerically closer between response variables, than the relative requirement estimates. However, the variability of the estimates precludes us from establishing statistical significance. By focusing on the requirements for rainbow trout, shows that none of the four mathematical models results in more precise estimates. As well, and contrary to the conclusions of Hauler and Carter (2001) or Bodin *et al.* (2009), we did not observe a significant difference in estimate deviation between the two relative (% diet dry matter and g/MJDE) and the absolute (g ingested/kg MBW/d) modes of

expression. However, the relationship between lysine requirement in rainbow trout and IBW depended on the unit of requirement (Figure 3): there is a significant linear regression when the requirement is expressed as g/kg MBW/d ( $P=0.0041$ ,  $R^2=0.482$ ), but not when express as percent of diet dry matter ( $P=0.3170$ ,  $R^2=0.077$ ). Therefore, differences in fish size account for some of the observed deviation, and can be better quantified using an absolute, intake-based mode of expression. Overall, the variability in requirement estimates limits our interpretation of EAA requirements in fish. It also underscores the need to better identify factors governing EAA requirements. This is clearly reflected by the debate over the mode of expression of EAA requirement. The three modes of expression chosen for this meta-analysis represent three, fundamentally different approaches which are based on mutually-exclusive assumptions. Therefore, there is a great need to further investigate the determinants of EAA requirements, and notably their dependence to dietary ingredients or feed intake.

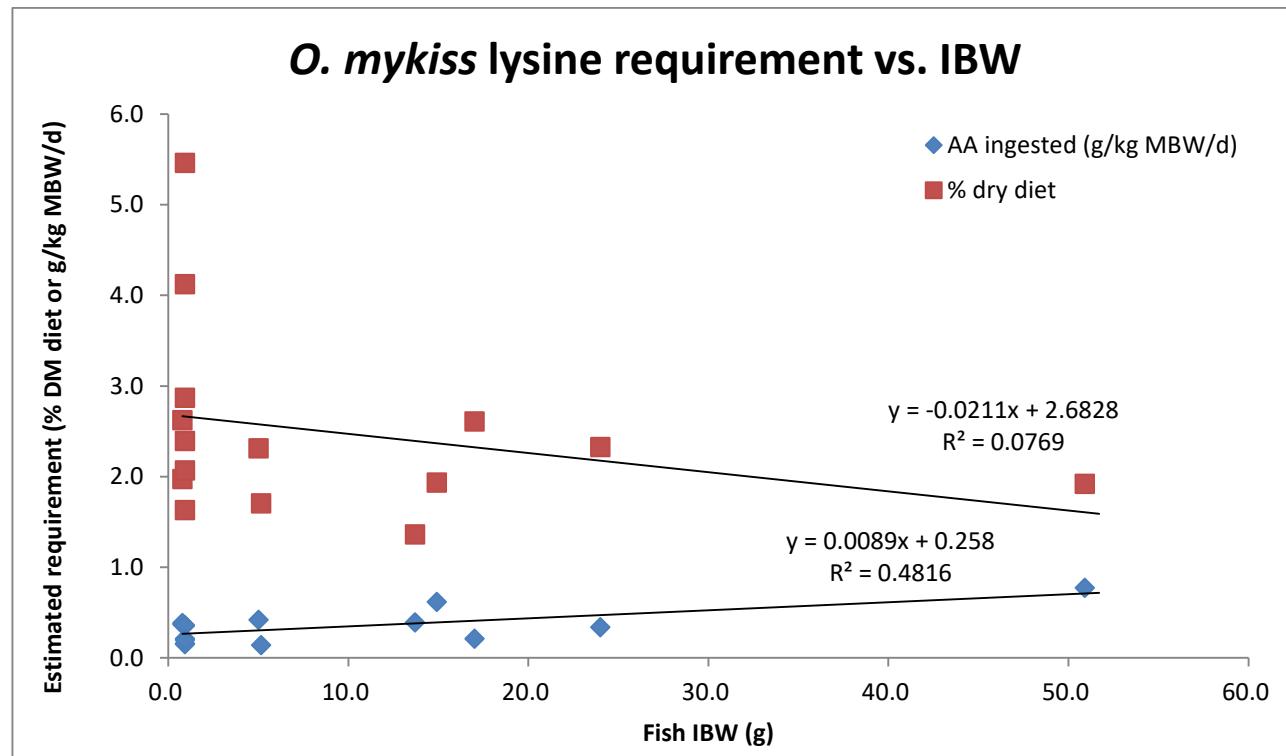


Figure 3: Relationship between initial body weight (IBW, g) and lysine requirement of *O. mykiss* expressed as a percent of diet dry matter or as an ingested amount.

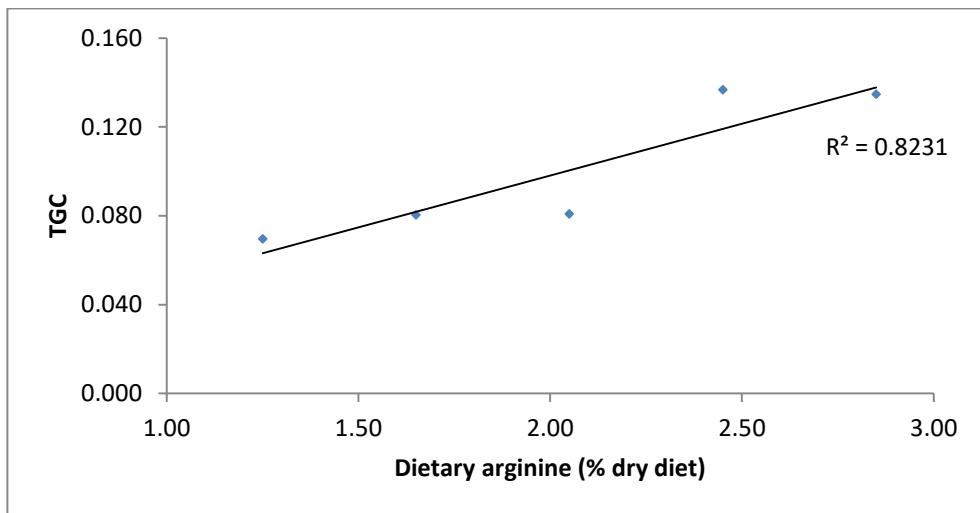


Figure 4: Growth response of turbot (*Psetta maxima*) to graded levels of dietary arginine, illustrating a linear pattern. Adapted from Fournier *et al.* (2003).

### 3.3 Overall limitations in the dataset

One of the biggest missing pieces of information in the present dataset is digestibility. Indeed, only 15 of the 145 selected studies measured and reported protein digestibility, and none measured the digestibility of individual EAA. Additionally, experimental diets were formulated using a wide range of ingredients. In the selected studies, diets were typically formulated using a mix of natural proteins (fishmeal, soy and corn proteins, etc.) and crystal amino acids. Not only the EAA supplied by these ingredients likely have different digestibility, but there may be relevant interactions among amino acids as well as with other non-protein components. These interactions may affect the availability of EAA to the animal, hence the apparent EAA requirement. For example, phytate is known to bind to amino acids – especially basic amino acids such as lysine, histidine and arginine – which limits their bioavailability to the animal (Chowdhury *et al.*, 2012; Selle *et al.*, 2006; Storebakken *et al.*, 2000). Therefore, data from studies using phytate-rich ingredients such as plant proteins should be carefully interpreted.

Table 3: Estimates of essential amino acid requirements computed using the quadratic model (QM)

Estimates of requirements					
Unit of requirement estimate	Percent of diet dry matter		g/MJ DE		g ingested AA/kg MBW/d
Growth variable	g/kg MBW/d	TGC	g/kg MBW/d	TGC	g/kg MBW/d
<b>Arg</b>					
<i>O. kisutch</i>	1.83 ± 69.9%	1.94 ± 60.9%	1.17 ± 68.5%	1.24 ± 59.8%	0.09 ± 8.5%    0.10 ± 1.2%
<i>O. mykiss</i>	1.66 ± 19.0%	1.69 ± 19.0%	0.95 ± 15.5%	0.97 ± 15.4%	0.23 ± 13.0%    0.23 ± 13.3%
<i>S. salar</i>	1.86 ± 4.8%	1.87 ± 3.5%	0.93 ± 10.3%	0.94 ± 11.6 %	0.11 ± 51.4%    0.11 ± 52.9%
<b>Ile</b>					
<i>O. tshawytscha</i>	0.97 ± 6.3%	0.98 ± 7.0%	0.65 ± 6.2%	0.66 ± 7.0%	0.02 ± 3.7%    0.02 ± 4.8%
<b>Lys</b>					
<i>I. punctatus</i>	1.53 ± 0.8%	1.55 ± 14.9%	1.44 ± 1.0%	1.46 ± 6.3%	0.35 ± 15.1%    0.36 ± 6.1%
<i>O. mykiss</i>	2.02 ± 24.0%	2.04 ± 27.9%	1.02 ± 25.7%	1.03 ± 36.6%	0.25 ± 26.9%    0.25 ± 36.4%
<i>S. salar</i>	1.95 ± 46.9%	1.96 ± 49.7%	0.97 ± 39.2%	0.98 ± 41.1%	0.10 ± 51.4%    0.11 ± 52.1%
<b>Met</b>					
<i>Hybrid striped bass</i>	0.96 ± 21.1%	0.97 ± 21.1%	0.75 ± 21.1%	0.76 ± 21.5%	0.24 ± 52.8%    0.25 ± 54.2%
<i>I. punctatus</i>	0.76 ± 21.1%	0.69 ± 40.0%	0.74 ± 17.7%	0.66 ± 36.4%	0.13 ± 46.1%    0.12 ± 61.4%
<i>O. mykiss</i>	0.61 ± 10.5%	0.63 ± 11.4%	0.37 ± 5.2%	0.37 ± 4.9%	0.09 ± 11.9%    0.09 ± 11.4%
<b>Thr</b>					
<i>Hybrid striped bass</i>	0.92 ± 0.5%	0.94 ± 0.0%	0.72 ± 0.8%	0.74 ± 1.0%	0.08 ± 9.6%    0.08 ± 9.0%
<i>O. mykiss</i>	1.26 ± 14.0%	1.27 ± 14.0%	0.70 ± 6.4%	0.70 ± 6.1%	0.16 ± 20.5%    0.16 ± 20.6%
<b>Trp</b>					
<i>O. mykiss</i>	0.29 ± 16.4%	0.29 ± 16.2%	0.17 ± 11.3%	0.17 ± 11.1%	0.03 ± 30.7%    0.03 ± 31.1%

Values are estimates of requirement in the indicated unit, ± the coefficient of variation.

Table 4: Comparisons of lysine requirement estimates in rainbow trout *O. mykiss*, depending on choice of variables and mathematical model

Estimates of requirements ( $\pm$ coefficient of variation)										
Unit of requirement estimate	Percent of diet dry matter			g/MJ DE			g ingested AA/kg MBW/d			
Growth variable	g/kg MBW/d	TGC	g/kg MBW/d	TGC	g/kg MBW/d	TGC	g/kg MBW/d	TGC		
<b>Broken line</b>	1.66	$\pm$ 1.73	$\pm$	0.81	$\pm$ 0.94	$\pm$	0.19	$\pm$ 0.18	$\pm$	
	21.6%	22.5%		21.4%	43.5%		30.9%	42.0%		
	(n=14)	(n=12)		(n=13)	(n=9)		(n=14)	(n=12)		
<b>Broken quadratic</b>	1.69	$\pm$ 1.72	$\pm$	0.86	$\pm$ 0.88	$\pm$	0.20	$\pm$ 0.21	$\pm$	
	17.1%	20.2%		17.6%	33.4%		19.7%	33.2%		
	(n=11)	(n=11)		(n=11)	(n=11)		(n=11)	(n=11)		
<b>Quadratic</b>	1.83	$\pm$ 1.86	$\pm$	0.96	$\pm$ 0.97	$\pm$	0.22	$\pm$ 0.22	$\pm$	
	22.9%	28.0%		22.8%	38.3%		28.3%	38.0%		
	(n=12)	(n=12)		(n=12)	(n=12)		(n=13)	(n=13)		
<b>Saturation kinetic</b>	1.99±	2.10	$\pm$	1.05	$\pm$ 1.13	$\pm$	0.28	$\pm$ 0.30	$\pm$	
	17.9%	12.4%		20.5%	22.2%		15.5%	24.1%		
	(n=8)	(n=6)		(n=7)	(n=6)		(n=5)	(n=6)		

The starting body weight of fish also greatly varies between studies, from 0.16g to 662g, although 60% of the studies started with fish between 0.5 and 25g. However, nutrient utilization is known to change with fish size, including in early juvenile stages (Dumas *et al.* 2007). Therefore, considering EAA requirements across growth stanza likely introduces a source of variation in the requirement estimates.

Finally, only a minority of studies reported final proximate analysis of the fish, or protein deposition results. This precluded us from including variables such as protein retention in our analysis, which have also been debated as a possible way to express EAA requirements.

### **3.4 Choice of mathematical model and experimental design**

Although the choice of mathematical model does not improve the variability of requirement estimates, there were significant differences in performances and quality of fit. The BLM, BQM and QM always converged, while the SKM failed to do so in 19.9% of the selected studies, all variable pairs combined. It should be noted here that a converged model does not sufficient to produce an accurate and/or precise estimation of the requirement. Table 5 shows which parameters of experimental design the models are most sensitive to, and the quality of the outcome beyond model convergence. Both BLM and SKM are sensitive to the number of experimental diets. For instance, each additional diet in the experimental design will make the BLM 0.23 times as likely to produce a minimum instead of a maximum (or 4.35 times as likely to produce a maximum,  $p=0.0028$ ). The SKM model, although closer to biological reality, demands careful experimental design. Although a low number of diets will not significantly increase the risk of failed fit, each additional diet will make the SKM 0.89 times as likely to result in an estimate outside the experimental range ( $p=0.0059$ ). This is problematic, and an “outside” fit should not be considered accurate.

Although fish growth was used as a screening criteria for constituting the working dataset, the maxFBW/IBW ratio still significantly affected how the QM and BQM models fitted the data (Table 5). High weight gain in the best (presumably replete) treatments during the experiment resulted in a reduced risk of producing an estimate outside the experimental range – odds ratio of 0.63 ( $p=0.0111$ ) and 0.71 ( $p=0.0084$ ) for the BQM and QM, respectively. In other words, sufficient weight gain during the experiment contributes to estimates within the experimental range, and in that sense, contribute to the accuracy of the estimates. More surprisingly, the BQM also has a significant tendency to fit a minimum (i.e., negative slope of the quadratic segment) at high maxFBW/IBW

values. Upon closer inspection of such studies, fish growth response to the graded EAA levels is always quite linear, as illustrated in Figure 4. However, all models are based on the law of “diminishing returns”, and thus are best suited for situations that illustrate such a pattern. In 50% of failed BQM cases (and 97% of failed BLM), the model could not fit one of the 2 lines. This highlights that models are sensitive to the presence of both the initial ascending portion of the curve, and the following plateau. Models’ sensitivity to the former is well illustrated by the ratio between the lowest and highest fish weight at the end of the study (minFBW/maxFBW).

Table 5: Odds ratio of maxFBW/IBW, minFBW/maxFBW, and number of diets affecting the fit outcome of the requirement models.

Parameter Outcome	Broken Model	Quadratic	Broken-Line Model	Quadratic Model	Saturation Kinetic Model
<b>maxFBW / IBW ratio</b>					
<b>Failed fit</b>	NS		NS	NS	NS
<b>Minimum</b>	1.15		NS	NS	
<b>No break point</b>	NS		NS		
<b>Outside estimate</b>	0.63		NS	0.71	NS
<b>minFBW / maxFBW ratio</b>					
<b>Failed fit</b>	NS		NS	NS	30.73
<b>Minimum</b>	NS		83.75	NS	
<b>No break point</b>	106.40		26.28		
<b>Outside estimate</b>	NS		0.31	17.71	3.20
<b>Number of Diets</b>					
<b>Failed fit</b>	NS		NS	NS	NS
<b>Minimum</b>	NS		0.23	NS	
<b>No break point</b>	NS		NS		
<b>Outside estimate</b>	NS		NS	NS	0.89

Note: values odds ratio of an outcome occurring vs. a converged fit resulting in an estimate within the experimental range. Minimum: the model has a minimum instead of a maximum; No break point: the breaking point could not be

determined; Failed fit: the model failed to converge; Outside estimate: the estimated requirement is outside of experimental range. NS: odd ratio non-significant different from 1; Empty cells indicate that this outcome is non-applicable to the model.

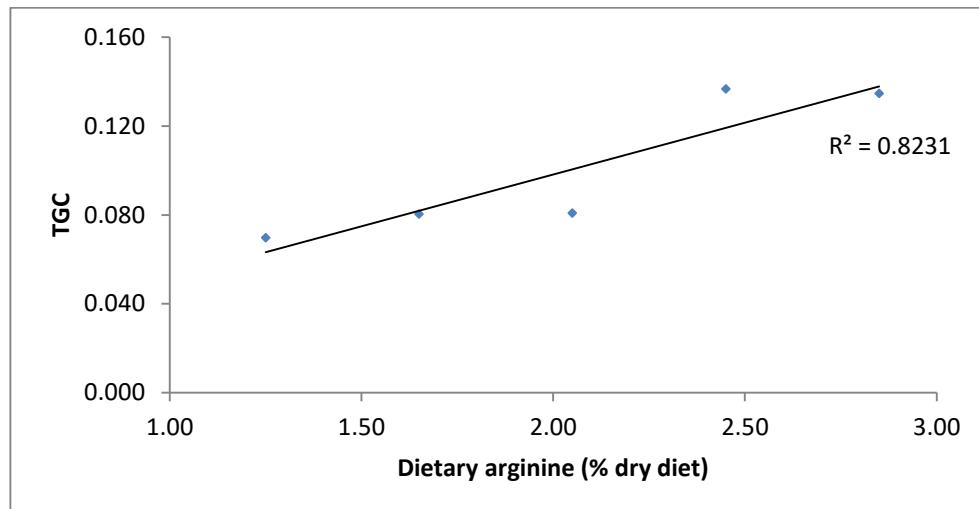


Figure 5: Growth response of turbot (*Psetta maxima*) to graded levels of dietary arginine, illustrating a linear pattern. Adapted from Fournier *et al.* (2003).

The minFBW/maxFBW was by far the most impactful metric regarding fit performance and quality in all models. If one could imagine a 1 unit increase in minFBW/maxFBW ratio, the BQM and the BLM would be 106.40 ( $p=0.0307$ ) and 26.28 ( $p=0.0319$ ) times as likely to fail in finding a breaking point, respectively. The BLM would also be 83.75 times as likely to fit a first line with a negative slope ( $p=0.0392$ ). Finally, the SKM would be 3.20 ( $p=0.0170$ ) and 30.73 ( $p<0.0001$ ) times as likely to find an estimate outside the experimental range and fail altogether, respectively. Evidently, studies failing to maximize the difference in final average weights between treatments at the end of the experiment are in great risk of not being able to accurately estimate the requirement. This occurs when the basal diet is not deficient enough and/or amino acid contents in other experimental diets

do not reach a high enough level to produce a clear plateau. Consequently, particular attention should be given to diet formulations to create such a pattern.

The above discussion highlights factors which nutritionists should carefully consider when developing the experimental design and formulating experimental diets. Our results show that none of the four models result in a significantly improved precision of the requirement estimate. The BLM has been a model of choice for decades, due to its simplicity and ease of fit, especially at a time when computers were not as ubiquitous. However, the limitations of the BLM have been brought to light (Hernandez-Llamas, 2009) and other models have since been suggested. However, no model can be *a priori* assumed to be superior in all situations, and a careful model selection procedure should be implemented for each experiment as part of the statistical evaluation of the results. Several methods exist for this, although the Akaike Information Criterion (AIC) method (Akaike, 1974; Arnold, 2010), which balances goodness of fit and parsimony of the models, is practical and provides guidance to the nutritionist in his/her choice of model selection (Salze *et al.* 2017).

#### **4 Conclusions and recommendations**

The standardization of the current body of scientific literature, as presently conducted, did not lead to clearer, more precise requirement estimates. However, it did highlight some important points. First, the quality of published studies is highly variable: less than half of the relevant studies were deemed suitable for our final data-set. The main reasons for these rejections include poor growth and a lack of reported information such as diet moisture content.

Future investigations of EAA requirements should focus on non-salmonid species, different life stages, and less-studied EAA such as taurine, phenylalanine, leucine, or histidine in order to remedy the knowledge fragmentation. Trials should use no fewer than 6 experimental diets, more if available facilities and resources allow. The use of the BLM should be avoided, as it has now been clearly demonstrated that it underestimates the requirement. Choice of mathematical model should then follow a rational evaluation of fit quality. SKM, QM, or BQM are all valid models, and none should

be *a priori* assumed inherently superior to another. In addition, it is important that the number and range of graded dietary levels of the test EAA are chosen so that the ascending and plateau segments of the curve can be clearly established with the requirement falling approximately in the middle. Finally, the effective loss of knowledge due to unreported information is very counterproductive. It is therefore most critical that fish nutritionists agree on a standard and systematic system to report and capture results of such trial.

## References

- Abidi, S.F. and Khan, M.A., 2004. Dietary valine requirement of Indian major carp, *Labeo rohita* (Hamilton) fry. Journal of Applied Ichthyology 20, 118–122.
- Abidi, S.F. and Khan, M.A., 2007. Dietary leucine requirement of fingerling Indian major carp, *Labeo rohita* (Hamilton). Aquaculture Research 38, 478-486.
- Abidi, S.F. and Khan, M.A., 2008. Dietary threonine requirement of fingerling Indian major carp, *Labeo rohita* (Hamilton). Aquaculture Research 39, 1498-1505.
- Abidi, S.F. and Khan, M.A., 2009. Dietary arginine requirement of fingerling Indian major carp, *Labeo rohita* (Hamilton) based on growth, nutrient retention efficiencies, RNA/DNA ratio and body composition. Journal of Applied Ichthyology 25 (6), 707-714.
- Abidi, S.F. and Khan, M.A., 2010. Dietary Tryptophan Requirement of Fingerling Rohu, *Labeo rohita* (Hamilton), Based on Growth and Body Composition. Journal of the World Aquaculture Society 41 (5), 700-709.
- Abimorad, E.G., Favero, G.C., Squassoni, G.H., Carneiro, D., 2010. Dietary digestible lysine requirement and essential amino acid to lysine ratio for pacu *Piaractus mesopotamicus*. Aquaculture Nutrition 16 (4), 370-377.
- Ahmed, I. and Khan, M.A., 2004a. Dietary lysine requirement of fingerling Indian major carp, *Cirrhinus mrigala* (Hamilton). Aquaculture 235, 499– 511.
- Ahmed, I. and Khan, M.A., 2004b. Dietary arginine requirement of fingerling Indian major carp, *Cirrhinus mrigala* (Hamilton). Aquaculture Nutrition 10, 217–225.
- Ahmed, I., Khan, M.A., Jafri, A.K., 2004. Dietary threonine requirement of fingerling Indian major carp, *Cirrhinus mrigala* (Hamilton). Aquaculture Research 35, 162-170.
- Ahmed, I. and Khan, M.A., 2005a. Dietary tryptophan requirement of fingerling Indian major carp, *Cirrhinus mrigala* (Hamilton). Aquaculture Research 36, 687-695.
- Ahmed, I. and Khan, M.A., 2005b. Dietary histidine requirement of fingerling Indian major carp, *Cirrhinus mrigala* (Hamilton). Aquaculture Nutrition 11, 359–366.
- Ahmed, I. and Khan, M.A., 2006. Dietary branched-chain amino acid valine, isoleucine and leucine requirements of fingerling Indian major carp, *Cirrhinus mrigala* (Hamilton). British Journal of Nutrition 96, 450-460.
- Ahmed, I., 2009. Dietary total aromatic amino acid requirement and tyrosine replacement value for phenylalanine in Indian major carp: *Cirrhinus mrigala* (Hamilton) fingerlings. Journal of Applied Ichthyology 25 (6), 719-727.
- Ahmed, I., 2012. Dietary amino acid L-tryptophan requirement of fingerling Indian catfish, *Heteropneustes fossilis* (Bloch), estimated by growth and haemato-biochemical parameters. Fish Physiology and Biochemistry 38 (4), 1195-1209.

- Ahmed, I., 2013. Dietary arginine requirement of fingerling Indian catfish (*Heteropneustes fossilis*, Bloch). *Aquaculture International* 21 (2), 255-271.
- Akaike, H., 1974. A new look at the statistical model identification. *IEEE Transactions on Automatic Control* 19 (6), 716-723.
- Akiyama, T., Arai, S., Murai, T., Nose, T., 1985. Threonine, Histidine and Lysine Requirements of Chum Salmon Fry. *Bulletin of the Japanese Society of Scientific Fisheries* 51 (4), 635-639.
- Alam, M.S., Teshima, S.-I., Ishikawa, M., Koshio, S., 2000. Methionine Requirement of Juvenile Japanese Flounder *Paralichthys olivaceus*. *Journal of the World Aquaculture Society* 31 (4).
- Alam, M.S., Teshima, S.-I., Ishikawa, M., Koshio, S., 2002. Effects of dietary arginine and lysine levels on growth performance and biochemical parameters of juvenile Japanese flounder *Paralichthys olivaceus*. *Fisheries Science* 68, 509–516.
- Anderson, J.S., Lall, S.P., Anderson, D.M., McNiven, M.A., 1993. Quantitative dietary lysine requirement of Atlantic salmon (*Salmo salar*) fingerlings. *Canadian Journal of Fisheries & Aquatic Sciences* 50 (2), 316-322.
- Arnold, T.W., 2010. Uninformative Parameters and Model Selection Using Akaike's Information Criterion. *Journal of Wildlife Management* 74 (6), 1175-1178.
- Berge, G.E., Lied, E., Sveier, H., 1997. Nutrition of Atlantic salmon (*Salmo salar*): The requirement and metabolism of arginine. *Comparative Biochemistry and Physiology - Part A: Molecular & Integrative Physiology* 117 (4), 501-509.
- Bodin, N., Mambrini, M., Wauters, J.-B., Abboudi, T., Ooghe, W., Le Boulenge, E., Larondelle, Y., Rollin, X., 2008. Threonine requirements for rainbow trout (*Oncorhynchus mykiss*) and Atlantic salmon (*Salmo salar*) at the fry stage are similar. *Aquaculture* 274 (2-4), 353-365.
- Bodin, N., Govaerts, B., Abboudi, T., Detavernier, C., De Saeger, S., Larondelle, Y., Rollin, X., 2009. Protein level affects the relative lysine requirement of growing rainbow trout (*Oncorhynchus mykiss*) fry. *British Journal of Nutrition* 102 (1), 37-53.
- Boren, R.S. and Gatlin III, D.M., 1995. Dietary Threonine Requirement of Juvenile Red Drum *Sciaenops ocellatus*. *Journal of the World Aquaculture Society* 26 (3), 279-283.
- Bureau, D.P. and Encarnaçāo, P., 2006. Adequately defining the amino acid requirements of fish: The case example of lysine. *Advances en Nutrición Acuícola VIII - VIII Simposium Internacional de Nutrición Acuícola*, Universidad Autónoma de Nuevo León, Monterrey, Nuevo León, Mexico, pp. 29-54.
- Bureau, D.P., 2008. Toward a better definition of essential amino acid requirements of fish, International Aquafeed. Perendale Publishers Ltd, pp. 14-17.
- Cai, Y. and Burtle, G.J., 1996. Methionine requirement of channel catfish fed soybean meal-corn-based diets. *J. Anim Sci.* 74 (3), 514-521.

- Cao, J.M., Chen, Y., Zhu, X., Huang, Y.H., Zhao, H.X., Li, G., Lan, H.B., Chen, B., Pan, Q., 2012. A study on dietary l-lysine requirement of juvenile yellow catfish *Pelteobagrus fulvidraco*. *Aquaculture Nutrition* 18 (1), 35-45.
- Chance, R.E., Mertz, E.T., Halver, J.E., 1964. Nutrition of Salmonoid Fishes: XII. Isoleucine, Leucine, Valine and Phenylalanine Requirements of Chinook Salmon and Interrelations between Isoleucine and Leucine for Growth. *Journal of Nutrition* 83 (3), 177-185.
- Cheng, Z.J., Hardy, R.W., Usry, J.L., 2003. Effects of lysine supplementation in plant protein-based diets on the performance of rainbow trout (*Oncorhynchus mykiss*) and apparent digestibility coefficients of nutrients. *Aquaculture* 215, 255–265.
- Cho, C.Y., Kaushik, S.J., Woodward, B., 1992. Dietary arginine requirement of young rainbow trout (*Oncorhynchus mykiss*). *Comparative Biochemistry and Physiology - Part A: Molecular & Integrative Physiology* 102 (1), 221-216.
- Chowdhury, M.A.K., Tacon, A.G.J., Bureau, D.P., 2012. Digestibility of amino acids in Indian mustard protein concentrate and Indian mustard meal compared to that of a soy protein concentrate in rainbow trout and Atlantic salmon. *Aquaculture* 356-357, 128-134.
- Chu, Z.J., Gong, Y., Lin, Y.C., Yuan, Y.C., Cai, W.J., Gong, S.Y., Luo, Z., 2014. Optimal dietary methionine requirement of juvenile Chinese sucker, *Myxocyprinus asiaticus*. *Aquaculture Nutrition* 20 (3), 253-264.
- Coloso, R.M., Murillo-Gurrea, D.P., Borlongan, I.G., Catacutan, M.R., 1999. Sulphur amino acid requirement of juvenile Asian sea bass *Lates calcarifer*. *Journal of Applied Ichthyology* 15, 54-58.
- Cowey, C.B., Cho, C.Y., Sivak, J.G., Weerheim, J.A., Stuart, D.D., 1992. Methionine Intake in Rainbow Trout (*Oncorhynchus mykiss*), Relationship to Cataract Formation and the Metabolism of Methionine. *Journal of Nutrition* 122 (5), 1154-1163.
- Cowey, C.B., 1994. Amino acid requirements of fish: a critical appraisal of present values. *Aquaculture* 124 (1-4), 1-11.
- Craig, S.R. and Gatlin, D.M., III, 1992. Dietary Lysine Requirement of Juvenile Red Drum *Sciaenops ocellatus*. *Journal of the World Aquaculture Society* 23 (2), 133-137.
- Dumas, A., France, J., Bureau, D.P., 2007. Evidence of three growth stanzas in rainbow trout (*Oncorhynchus mykiss*) across life stages and adaptation of the thermal-unit growth coefficient. *Aquaculture* 267 (1-4), 139-146.
- Encarnacao, P., Langea, C.d., Rodehutscordb, M., Hoehlrc, D., Bureaua, W., Bureau, D.P., 2004. Diet digestible energy content affects lysine utilization, but not dietary lysine requirements of rainbow trout (*Oncorhynchus mykiss*) for maximum growth. *Aquaculture* 235 569–586.
- Espe, M., Lemme, A., Petri, A., El-Mowafy, A., 2007. Assessment of lysine requirement for maximal protein accretion in Atlantic salmon using plant protein diets. *Aquaculture* 263 (1-4), 168-178.

- Espe, M., Hevrøy, E.M., Liaset, B., Lemme, A., El-Mowafi, A., 2008. Methionine intake affect hepatic sulphur metabolism in Atlantic salmon, *Salmo salar*. Aquaculture 274 (1), 132-141.
- Farhatand Khan, M.A., 2014. Dietary L-tryptophan requirement of fingerling stinging catfish, *Heteropneustes fossilis* (Bloch). Aquaculture Research 45 (7), 1224-1235.
- Forster, I. and Ogata, H.Y., 1998. Lysine requirement of juvenile Japanese flounder *Paralichthys olivaceus* and juvenile red sea bream *Pagrus major*. Aquaculture 161 (1-4), 131-142.
- Fournier, V., Gouillou-Coustans, M.F., Métailler, R., Vachot, C., Moriceau, J., Le Delliou, H., Huelvan, C., Desbruyeres, E., Kaushik, S.J., 2003. Excess dietary arginine affects urea excretion but does not improve N utilisation in rainbow trout *Oncorhynchus mykiss* and turbot *Psetta maxima*. Aquaculture 217 (1-4), 559-576.
- Furuya, W.M., Hayashi, C., Furuya, V.R.B., Botaro, D., Silva, L.C.d., Neves, P.R., 2008. Exigências de metionina + cistina total e digestível para alevinos revertidos de tilápia do Nilo, *Oreochromis niloticus* (L.), baseadas no conceito de proteína ideal. Acta Scientiarum. Animal Sciences 23 (4), 885-889.
- Gan, L., Zhou, L.L., Li, X.X., Yue, Y.R., 2016. Dietary leucine requirement of Juvenile Nile tilapia, *Oreochromis niloticus*. Aquaculture Nutrition 22 (5), 1040-1046.
- Gao, Y.J., Yang, H.J., Guo, D.Q., Liu, Y.J., Yu, Y.Y., Chen, S.J., Tian, L.X., 2015. Dietary arginine requirement of juvenile grass carp *Ctenopharyngodon idella* (Valenciennes in Cuvier & Valenciennes, 1844). Aquaculture Research 46 (12), 3070-3078.
- Griffin, M.E., Brown, P.B., Grant, A.L., 1992. The Dietary Lysine Requirement of Juvenile Hybrid Striped Bass. Journal of Nutrition 122 (6), 1332-1337.
- Habte-Tsion, H.M., Liu, B., Ren, M.C., Ge, X.P., Xie, J., Zhou, Q.L., Miao, L.H., Pan, L.K., Chen, R.L., 2015. Dietary threonine requirement of juvenile blunt snout bream (*Megalobrama amblycephala*). Aquaculture 437, 304-311.
- Harding, D.E., Allen, O.W., Jr., Wilson, R.P., 1977. Sulfur Amino Acid Requirement of Channel Catfish: L-Methionine and L-Cystine. Journal of Nutrition 107 (11), 2031-2035.
- Hauler, R.C. and Carter, C.G., 2001. Reevaluation of the quantitative dietary lysine requirements of fish. Reviews in Fisheries Science 9 (3), 133.
- Hauler, R.C., Carter, C.G., Edwards, S.J., 2007. Feeding regime does not influence lysine utilisation by Atlantic salmon, *Salmo salar* L., parr. Aquaculture 273 (4), 545-555.
- He, Z.G., Mai, K.S., Li, Y., Cheng, Z.Y., Ai, Q.H., 2016. Dietary threonine requirement of juvenile large yellow croaker, *Larmichthys crocea*. Aquaculture Research 47 (11), 3616-3624.
- Hernandez-Llamas, A., 2009. Conventional and alternative dose-response models to estimate nutrient requirements of aquaculture species. Aquaculture 292 (3-4), 207-213.
- Hughes, S.G., Rumsey, G.L., Nesheim, M.C., 1983. Dietary requirements for essential branched-chain amino acids by lake trout. Transactions of the American Fisheries Society 112 (6), 812-817.

- Keembiyehetty, C.N.and Gatlin, D.M., 1992. Dietary lysine requirement of juvenile hybrid striped bass (*Morone chrysops* X *M. saxatilis*). *Aquaculture* 104 (3-4), 271-277.
- Keembiyehetty, C.N.and Gatlin, D.M., 1993. Total sulfur amino acid requirement of juvenile hybrid striped bass (*Morone chrysops*×*M. saxatilis*). *Aquaculture* 110 (3-4), 331-339.
- Keembiyehetty, C.N.and Gatlin, D.M., III, 1997. Dietary threonine requirement of juvenile hybrid striped bass (*Morone chrysops* ♀ x *M. saxatilis* ♂). *Aquaculture Nutrition* 3 (4), 217-221.
- Khan, M.A.and Abidi, S.F., 2007a. Total aromatic amino acid requirement of Indian major carp *Labeo rohita* (Hamilton) fry. *Aquaculture* 267 (1-4), 111-118.
- Khan, M.A.and Abidi, S.F., 2007b. Dietary isoleucine requirement of fingerling Indian major carp, *Labeo rohita* (Hamilton). *Aquaculture Nutrition* 13 (6), 424-430.
- Khan, M.A.and Abidi, S.F., 2014. Dietary histidine requirement of Singhi, *Heteropneustes fossilis* fry (Bloch). *Aquaculture Research* 45 (8), 1341-1354.
- Kim, K.-I., Kayes, T.B., Amundson, C.H., 1992a. Requirements for sulfur amino acids and utilization of D-methionine by rainbow trout (*Oncorhynchus mykiss*). *Aquaculture* 101 (1-2), 95-103.
- Kim, K.-I., Kayes, T.B., Amundson, C.H., 1992b. Requirements for lysine and arginine by rainbow trout (*Oncorhynchus mykiss*). *Aquaculture* 106 (3-4), 333-344.
- Kim, K.-I., 1993. Requirement for phenylalanine and replacement value of tyrosine for phenylalanine in rainbow trout (*Oncorhynchus mykiss*). *Aquaculture* 113 (3), 243-250.
- Klein, R.G.and Halver, J.E., 1970. Nutrition of Salmonoid Fishes: Arginine and Histidine Requirements of Chinook and Coho Salmon. *Journal of Nutrition* 100 (9), 1105-1109.
- Lall, S.P., Kaushik, S.J., Le Bail, P.Y., Keith, R., Anderson, J.S., Plisetskaya, E., 1994. Quantitative arginine requirement of Atlantic salmon (*Salmo salar*) reared in sea water. *Aquaculture* 124, 13-25.
- Lall, S.P.and Anderson, S., 2005. Amino acid nutrition of salmonids: Dietary requirements and bioavailability. *Cahiers Options Méditerranéennes* 63, 73-90.
- Li, Y., Cheng, Z.Y., Mai, K.S., Ai, Q.H., 2015. Dietary leucine requirement of juvenile Japanese seabass (*Lateolabrax japonicus*). *Journal of Ocean University of China* 14 (1), 121-126.
- Liao, Y.J., Ren, M.C., Liu, B., Sun, S.M., Cui, H.H., Xie, J., Zhou, Q.L., Pan, L.K., Chen, R.L., Ge, X.P., 2014. Dietary methionine requirement of juvenile blunt snout bream (*Megalobrama amblycephala*) at a constant dietary cystine level. *Aquaculture Nutrition* 20 (6), 741-752.
- Luzzana, U., Hardy, R.W., Halver, J.E., 1998. Dietary arginine requirement of fingerling coho salmon (*Oncorhynchus kisutch*). *Aquaculture* 163 (1-2), 137-150.

- Masagounder, K., Hayward, R.S., Firman, J.D., 2011. Comparison of dietary essential amino acid requirements determined from group-housed versus individually-housed juvenile bluegill, *Lepomis macrochirus*. *Aquaculture Nutrition* 17 (2), E559-E571.
- Michelato, M., Vidal, L.V.O., Xavier, T.O., Graciano, T.S., De Moura, L.B., Furuya, V.R.B., Furuya, W.M., 2016. Dietary threonine requirement to optimize protein retention and fillet production of fast-growing Nile tilapia. *Aquaculture Nutrition* 22 (4), 759-766.
- Moon, H.Y. and Gatlin III, D.M., 1991. Total sulfur amino acid requirement of juvenile red drum, *Sciaenops ocellatus*. *Aquaculture* 95 (1-2), 97-106.
- Murillo-Gurrea, D.P., Coloso, R.M., Borlongan, I.G., Serrano, A.E., 2001. Lysine and arginine requirements of juvenile Asian sea bass (*Lates calcarifer*). *Journal of Applied Ichthyology* 17 (2), 49-53.
- Nang Thu, T.T., Parkouda, C., Saeger, S.d., Larondelle, Y., Rollin, X., 2007. Comparison of the lysine utilization efficiency in different plant protein sources supplemented with l-lysine·HCl in rainbow trout (*Oncorhynchus mykiss*) fry. *Aquaculture* 272 (1-4), 477-488.
- Nang Thu, T.T., Parkouda, C., de Saeger, S., Larondelle, Y., Rollin, X., 2009. Protein level does not affect lysine utilization efficiency at marginal lysine intake in growing rainbow trout (*Oncorhynchus mykiss*) fry. *Aquaculture* 288 (3-4), 312-320.
- Niu, J., Du, Q., Lin, H.Z., Cheng, Y.Q., Huang, Z., Wang, Y., Wang, J., Chen, Y.F., 2013. Quantitative dietary methionine requirement of juvenile golden pompano *Trachinotus ovatus* at a constant dietary cystine level. *Aquaculture Nutrition* 19 (5), 677-686.
- NRC, 1993. Nutrient Requirements of Fish. National Acad. Press, Washington, DC.
- NRC, 2011. Nutrient requirements of fish and shrimp. The National Academies Press, Washington, D.C., 360 pp.
- Peres, H. and Oliva-Teles, A., 2008. Lysine requirement and efficiency of lysine utilization in turbot (*Scophthalmus maximus*) juveniles. *Aquaculture* 275 (1-4), 283-290.
- Pfeffer, E., Al-Sabty, H., Haverkamp, R., 1992. Studies on lysine requirements of rainbow trout (*Oncorhynchus mykiss*) fed wheat gluten as only source of dietary protein. *Journal of Animal Physiology and Animal Nutrition* 67 (1-5), 74-82.
- Poston, H.A. and Rumsey, G.L., 1983. Factors Affecting Dietary Requirement and Deficiency Signs of L-Tryptophan in Rainbow Trout. *Journal of Nutrition* 113 (12), 2568-2577.
- Ren, M.C., Ai, Q.H., Mai, K.S., 2014. Dietary arginine requirement of juvenile cobia (*Rachycentron canadum*). *Aquaculture Research* 45 (2), 225-233.
- Robinson, E.H., Wilson, R.P., Poe, W.E., 1980a. Total Aromatic Amino Acid Requirement, Phenylalanine Requirement and Tyrosine Replacement Value for Fingerling Channel Catfish. *Journal of Nutrition* 110 (9), 1805-1812.

- Robinson, E.H., Wilson, R.P., Poe, W.E., 1980b. Re-evaluation of the Lysine Requirement and Lysine Utilization by Fingerling Channel Catfish. *Journal of Nutrition* 110 (11), 2313-2316.
- Robinson, E.H., Wilson, R.P., Poe, W.E., 1981. Arginine Requirement and Apparent Absence of a Lysine-Arginine Antagonist in Fingerling Channel Catfish. *Journal of Nutrition* 111 (1), 46-52.
- Rodehutscord, M., Jacobs, S., Pack, M., Pfeffer, E., 1995a. Response of Rainbow Trout (*Oncorhynchus mykiss*) Growing from 50 to 170 g to Supplements of Either L-Arginine or L-Threonine in a Semipurified Diet. *Journal of Nutrition* 125 (4), 970-975.
- Rodehutscord, M., Jacobs, S., Pack, M., Pfeffer, E., 1995b. Response of Rainbow Trout (*Oncorhynchus mykiss*) Growing from 50 to 150 g to Supplements of DL-Methionine in a Semipurified Diet Containing Low or High Levels of Cystine. *Journal of Nutrition* 125 (4), 964-969.
- Rodehutscord, M., Becker, A., Pack, M., Pfeffer, E., 1997. Response of Rainbow Trout (*Oncorhynchus mykiss*) to Supplements of Individual Essential Amino Acids in a Semipurified Diet, Including an Estimate of the Maintenance Requirement for Essential Amino Acids. *Journal of Nutrition* 127 (6), 1166-1175.
- Rollin, X., Wauters, J.B., Bodin, N., Larondelle, Y., Ooghe, W., Wathelet, B., Abboudi, T., 2006. Maintenance threonine requirement and efficiency of its use for accretion of whole-body threonine and protein in Atlantic salmon (*Salmo salar* L.) fry. *British Journal of Nutrition* 95 (2), 234-245.
- Salze, G.P., Stuart, K.R., Jirsa, D.O., Davis, D.A., Drawbridge, M.A., 2017. Quantitative Dietary Taurine Requirement for California Yellowtail, *Seriola lalandi*. *Journal of the World Aquaculture Society*.
- Selle, P.H., Ravindran, V., Bryden, W.L., Scott, T., 2006. Influence of dietary phytate and exogenous phytase on amino acid digestibility in poultry : A review. *The Journal of Poultry Science* 43, 89-103.
- Shearer, K.D., 2000. Experimental design, statistical analysis and modelling of dietary nutrient requirement studies for fish: a critical review. *Aquaculture Nutrition* 6 (2), 91-102.
- Simmons, L., Moccia, R.D., Bureau, D.P., Sivak, J.G., Herbert, K., 1999. Dietary methionine requirement of juvenile Arctic charr *Salvelinus alpinus* (L.). *Aquaculture Nutrition* 5 (2), 93-100.
- Singh, S. and Khan, M.A., 2007. Dietary arginine requirement of fingerling hybrid *Clarias* (*Clarias gariepinus* x *Clarias macrocephalus*). *Aquaculture Research* 38 (1), 17-25.
- Storebakken, T., Shearer, K.D., Baeverfjord, G., Nielsen, G., Åsgård, T., Scott, T., De Laporte, A., 2000. Digestibility of macronutrients, energy and amino acids, absorption of elements and absence of intestinal enteritis in Atlantic salmon, *Salmo salar*, fed diets with wheat gluten. *Aquaculture* 184, 115-132.
- Tibaldi, E. and Lanari, D., 1991. Optimal dietary lysine levels for growth and protein utilisation of fingerling sea bass (*Dicentrarchus labrax* L.) fed semipurified diets. *Aquaculture* 95 (3-4), 297-304.
- Tibaldi, E. and Tulli, F., 1999. Dietary threonine requirement of juvenile european sea bass (*Dicentrarchus labrax*). *Aquaculture* 175 (1-2), 155-166.

- Walton, M., Cowey, C., Coloso, R., Adron, J., 1986. Dietary requirements of rainbow trout for tryptophan, lysine and arginine determined by growth and biochemical measurements. *Fish Physiology and Biochemistry* 2 (1), 161-169.
- Walton, M.J., Coloso, R.M., Cowey, C.B., Adron, J.W., Knox, D., 1984a. The effects of dietary tryptophan levels on growth and metabolism of rainbow trout (*Salmo gairdneri*). *British Journal of Nutrition* 51 (02), 279-287.
- Walton, M.J., Cowey, C.B., Adron, J.W., 1984b. The effect of dietary lysine levels on growth and metabolism of rainbow trout (*Salmo gairdneri*). *British Journal of Nutrition* 52 (01), 115-122.
- Wang, S., Liu, Y.-J., Tian, L.-X., Xie, M.-Q., Yang, H.-J., Wang, Y., Liang, G.-Y., 2005. Quantitative dietary lysine requirement of juvenile grass carp *Ctenopharyngodon idella*. *Aquaculture* 249 (1-4), 419-429.
- Wang, S., Encarnaçao, P., Bureau, D.P., unpublished. Effects of dietary lysine level on growth performance, nutrient utilization, muscle cellularity and flesh quality of juvenile rainbow trout (*Oncorhynchus mykiss*).
- Wang, Y.Y., Che, J.F., Tang, B.B., Yu, S.L., Wang, Y.Y., Yang, Y.H., 2016. Dietary methionine requirement of juvenile *Pseudobagrus ussuriensis*. *Aquaculture Nutrition* 22 (6), 1293-1300.
- Wilson, R.P., Harding, D.E., Garling, D.L., Jr., 1977. Effect of Dietary pH on Amino Acid Utilization and the Lysine Requirement of Fingerling Channel Catfish. *Journal of Nutrition* 107 (1), 166-170.
- Wilson, R.P., Allen, O.W., Jr., Robinson, E.H., Poe, W.E., 1978. Tryptophan and Threonine Requirements of Fingerling Channel Catfish. *Journal of Nutrition* 108 (10), 1595-1599.
- Wilson, R.P., Poe, W.E., Robinson, E.H., 1980. Leucine, Isoleucine, Valine and Histidine Requirements of Fingerling Channel Catfish. *Journal of Nutrition* 110 (4), 627-633.
- Wilson, R.P., 1989. Protein and amino acid requirements of fishes. Progress in fish nutrition. Proceedings of the fish nutrition symposium, 9, pp. 51-77.
- Zehra, S.and Khan, M.A., 2013a. Dietary Arginine Requirement of Fingerling Indian Major Carp, Catla catla (Hamilton). *Journal of the World Aquaculture Society* 44 (3), 363-373.
- Zehra, S.and Khan, M.A., 2013b. Dietary lysine requirement of fingerling Catla catla (Hamilton) based on growth, protein deposition, lysine retention efficiency, RNA/DNA ratio and carcass composition. *Fish Physiology and Biochemistry* 39 (3), 503-512.
- Zehra, S.and Khan, M.A., 2013c. Dietary isoleucine requirement of fingerling catla, Catla catla (Hamilton), based on growth, protein productive value, isoleucine retention efficiency and carcass composition. *Aquaculture International* 21 (6), 1243-1259.
- Zehra, S.and Khan, M.A., 2014. Dietary phenylalanine requirement and tyrosine replacement value for phenylalanine for fingerling Catla catla (Hamilton). *Aquaculture* 433, 256-265.

- Zehra, S. and Khan, M.A., 2015. Dietary tryptophan requirement of fingerling Catla catla (Hamilton) based on growth, protein gain, RNA/DNA ratio, haematological parameters and carcass composition. *Aquaculture Nutrition* 21 (5), 690-701.
- Zehra, S. and Khan, M.A., 2016a. Dietary threonine requirement of fingerling Indian major carp, Catla catla (Hamilton) estimated by growth, protein retention efficiency, threonine deposition, haematological parameters and carcass composition. *Aquaculture Research* 47 (1), 253-265.
- Zehra, S. and Khan, M.A., 2016b. Total sulphur amino acid requirement and maximum cysteine replacement value for methionine for fingerling Catla catla (Hamilton). *Aquaculture Research* 47 (1), 304-317.
- Zehra, S. and Khan, M.A., 2016c. Dietary histidine requirement of fingerling Catla Catla (Hamilton) based on growth, protein gain, histidine gain, RNA/DNA ratio, haematological indices and carcass composition. *Aquaculture Research* 47 (4), 1028-1039.
- Zhou, F., Shao, J., Xu, R., Ma, J., Xu, Z., 2009. Quantitative L-lysine requirement of juvenile black sea bream (*Sparus macrocephalus*). *Aquaculture Nutrition* 16 (2), 194-204.
- Zhou, F., Shao, J., Xu, R., Ma, J., Xu, Z., 2010a. Quantitative l-lysine requirement of juvenile black sea bream (*Sparus macrocephalus*). *Aquaculture Nutrition* 16 (2), 194-204.
- Zhou, F., Xiong, W., Xiao, J.X., Shao, Q.J., Bergo, O.N., Hua, Y., Chai, X.J., 2010b. Optimum arginine requirement of juvenile black sea bream, *Sparus macrocephalus*. *Aquaculture Research* 41 (10), e418-e430.
- Zhou, F., Xiao, J.X., Hua, Y., Ngandzali, B.O., Shao, Q.J., 2011. Dietary L-methionine requirement of juvenile black sea bream (*Sparus macrocephalus*) at a constant dietary cystine level. *Aquaculture Nutrition* 17 (5), 469-481.
- Zhou, H., Chen, N., Qiu, X., Zhao, M., Jin, L., 2012. Arginine requirement and effect of arginine intake on immunity in largemouth bass, *Micropterus salmoides*. *Aquaculture Nutrition* 18 (1), 107-116.
- Zhou, Q.-C., Wu, Z.-H., Tan, B.-P., Chi, S.-Y., Yang, Q.-H., 2006. Optimal dietary methionine requirement for Juvenile Cobia (*Rachycentron canadum*). *Aquaculture* 258 (1-4), 551-557.
- Zhou, Q.-C., Wu, Z.-H., Chi, S.-Y., Yang, Q.-H., 2007. Dietary Lysine Requirement of Juvenile Cobia (*Rachycentron canadum*). *Aquaculture* 273, 634-640.